Relaxant effect of the H₂-receptor antagonist oxmetidine on guinea-pig and human airways

Charles Advenier, Jean Pierre Gnassounou & *,1 Carmelo Scarpignato

Laboratoire de Pharmacologie, Faculté Médecine, Paris-Ouest, 15, rue de l'Ecole de Medécine, 75270 Paris Cedex 06, France and *Institute of Pharmacology, Faculty of Medicine and Dentistry, University of Parma, 43100 Parma, Italy

- 1 The effects of three different H₂-receptor antagonists (cimetidine, ranitidine and oxmetidine) were tested on isolated preparations of guinea-pig trachea and human bronchus against contractions induced by acetylcholine, histamine and potassium chloride (KCl). In addition, their influence on calcium concentration-response curves in guinea-pig tracheal spirals was examined in a potassium-rich solution (30 mM). Finally, their effects were studied *in vivo* against acetylcholine and histamine-induced bronchoconstriction in anaesthetized guinea-pigs.
- 2 In guinea-pig isolated trachea, oxmetidine in contrast to cimetidine and ranitidine, which were completely inactive induced a concentration-dependent relaxation regardless of the excitatory stimulus: its $\log EC_{50}$ values (i.e. the negative log concentration that caused a 50% relaxation) were 3.46 ± 0.11 , 4.61 ± 0.09 and 4.20 ± 0.12 against acetylcholine, histamine and KCl, respectively. In Ca^{2+} -free, K⁺-enriched solution, the compound was able to inhibit Ca^{2+} -induced contractions at concentrations close to those needed to counteract the spasmogenic effect of histamine in normal Krebs solution. Results obtained in the human bronchus preparation were similar to those observed in guinea-pig tracheal spirals.
- 3 When tested against acetylcholine or histamine-induced bronchoconstriction in vivo, oxmetidine (10 and 30 mg Kg^{-1} intravenously) significantly reduced the increase in pulmonary airway resistance (R_{aw}) induced by both agents. Once again, cimetidine and ranitidine were completely ineffective.
- 4 In summary, oxmetidine displayed non-specific antispasmogenic activity on guinea-pig and human airways. This effect, which is independent of H_2 -receptor blockade, represents a side-effect of the drug which may be connected to its interference with Ca^{2+} influx and the action or release of intracellular Ca^{2+} .

Introduction

H₂-receptor antagonists represent a well-known class of antiulcer drugs, their established mechanism of action being the inhibition of gastric acid secretion through H₂-receptor blockade on parietal cells (Bertaccini & Dobrilla, 1980). This antisecretory effect is the consequence of their primary pharmacological action (i.e. competitive antagonism at H₂-receptors) and it is shared by all the members of this group of drugs. However, both experimental and clinical studies have shown that many of these compounds possess some pharmacological actions completely independent of H₂-receptor blockade. These 'side-effects' are peculiar to an individual member rather than the entire class of the drugs (Bertaccini & Coruzzi, 1984a). Ranitidine, a compound bearing an

oxmetidine (an imidazole compound which differs from cimetidine in carrying an isocytosine ring instead of the cyanoguanidine group in the side chain, Figure 1) is endowed, in contrast, with inhibitory

alkyl furan ring, can be used to illustrate this point. Ranitidine possesses, as a side-effect, a cholinergic-like action, first demonstrated on gastrointestinal motility (Bertaccini & Coruzzi, 1982; Bertaccini & Scarpignato, 1982) and later also demonstrated on salivary secretion in the rat and on the guinea-pig urinary bladder (for review see Bertaccini & Coruzzi, 1984b). This cholinomimetic effect is involved in the increase in lower oesophageal sphincter (LES) pressure (Bertaccini et al., 1981) and in the decrease in gastric emptying (Scarpignato et al., 1982) observed in man and may be, therefore, of clinical relevance.

A more recent member of this class of drugs,

'Author for correspondence at University of Parma.

Figure 1 Chemical structures of the H_2 -receptor antagonists studied.

Oxmetidine

activity on the motility of the digestive tract (Bertaccini et al., 1983). In addition, this compound has been shown to possess a negative inotropic effect on isolated heart preparations from rabbits (Coruzzi et al., 1982) and guinea-pigs (Black et al., 1985) and on human atria (Coruzzi et al., 1983). Also, it was able to antagonize the contractile effects of KCl, angiotensin and noradrenaline on rabbit isolated aorta (Bertaccini et al., 1984). Concentrations required for this effect were 10 to 100 times higher than those needed to inhibit histamine H₂-receptors.

The mechanism of the cholinomimetic action of ranitidine involves both direct and indirect (through inhibition of acetylcholinesterase and butyryl cholinesterase) effects on muscarinic receptors (Bertaccini & Coruzzi, 1984b). In contrast, the mechanism of the antispasmogenic action of oxmetidine is not completely understood. Calcium antagonist activity (Coruzzi et al., 1982) and effects on the availability of intracellular calcium (Bertaccini et al., 1984) were suggested to explain this peculiar pharmacological action.

In contrast to cimetidine and ranitidine, oxmetidine was able to modify bronchomotor tone after oral administration in healthy humans (Dal Negro et al., 1984). In this connection we tested oxmetidine, as well as ranitidine and cimetidine, for effects on guinea-pig airways both in vitro and in vivo. Additional

experiments were performed on the human isolated bronchus preparation.

Preliminary results of the present investigation have been presented to the French Society of Pharmacology and have appeared in abstract form (Gnassounou et al., 1986).

Methods

Guinea-pig trachea in vitro

Tracheal spirals were obtained from male guinea-pigs (250–300 g) anaesthetized with urethane (1.25 g kg⁻¹) and were equilibrated under an initial tension of 1.5 g in Krebs solution at 37°C, gassed with 95% O₂ and 5% CO₂. The initial tension ensured that, after 1 h 15 min equilibration, the resting tension ranged between 0.4 and 0.6 g. Responses to agonists were reproducible under these conditions. Tension was measured isometrically with a Ugo BASILE isometric transducer and recorded on a Ugo BASILE writing microdynamometer. The Krebs solution had the following composition (mM), NaCl 113, KCl 4.7, CaCl₂ 1.9, MgSO₄ 1.2, NaHCO₃ 25, KH₂PO₄ 1.2, glucose 11.5.

Preparations were first tested for maximal tension with acetylcholine (ACh, 3×10^{-4} M) and for maximal relaxation with isoprenaline (3×10^{-6} M). Afterwards, the effects of oxmetidine, ranitidine and cimetidine were studied on contractions induced by ACh (2×10^{-5} M), histamine (2×10^{-5} M) and KCl (1×10^{-2} M). These H₂-receptor antagonists were added to the bath 15 min before the addition of the contractile agent. Test tissues were exposed to only one spasmogen followed by increasing concentrations of the H₂-receptor antagonists.

The effects of the H₂-receptor antagonists were expressed as % reduction of the initial contraction obtained with the stimulatory compound alone. — log EC₅₀ values (i.e. the negative log of the drug concentration that caused a 50% reduction) were calculated from the log concentration-effect curves for each H₂-receptor antagonist.

Experiments in Ca2+-free, K+-enriched medium

Ca²⁺ concentration-response curves were established according to Godfraind *et al.* (1968). Tracheal spirals were incubated for 1 h in a Krebs solution similar to the one described above, but without CaCl₂ then for 15 min in CaCl₂-free solution in the presence of ethylenediaminetetraacetic acid (EDTA) 10⁻³ M. The preparations were washed at intervals of 15 min. Subsequently, the spirals were incubated in a calciumfree solution with additional K⁺. The composition of the potassium-enriched solution was (mM): NaCl 109,

KCl 30, MgCl₂ 0.49, NaHCO₃ 11.9, Na₂HPO₄ 0.4 and glucose 5.5. This medium was gassed with 95% O₂ plus 5% CO₂ and, after equilibration, its final pH was 7.46. After incubation, the concentration-response curves to Ca²⁺ 0.01 to 3 mM were constructed by cumulative addition. The contact time between the tissue and each Ca²⁺ concentration was 10 to 15 min. The H₂-receptor antagonists were added to the bath 20 min before the addition of Ca²⁺. Three Ca²⁺ concentration-response curves (without and with two different concentrations of an H₂-antagonist) were constructed on each tissue, so it acted as its own control.

Experiments on human isolated bronchus

The human isolated bronchus was prepared according to Advenier et al. (1986). Human bronchial tissue (usually with an inner diameter of 4-12 mm) obtained from patients undergoing surgery for lung cancer, but taken at distance from malignancy, was dissected free of parenchyma and transported to the laboratory in ice-cold Krebs solution previously gassed with a mixture of 95% O₂ and 5% CO₂. The tissue was stored overnight at 4°C and the experiments were carried out on the next day. Previous investigations have shown that overnight storage of tissue does not alter its reactivity (Brink et al., 1980; Ghelani et al., 1980; Guillot et al., 1984; Vincenc et al., 1984). Spirally-cut strips from a segmental bronchus were suspended in Krebs solution under an initial tension of 2.5 g in the conditions described for guinea-pig isolated trachea. They were first contracted to maximal tension with carbachol ($1 \times 10^{-4} \,\mathrm{M}$). The effect of oxmetidine was subsequently tested against contractions induced by histamine, acetylcholine or KCl (at the concentrations stated above). When a stable contraction was obtained, cumulative concentration-response curves to oxmetidine (10^{-6} to 3×10^{-4} M) were obtained by adding increasing concentrations of the drugs at 10-20 min intervals. After the last concentration of oxmetidine, theophylline $(3 \times 10^{-3} \,\mathrm{M})$ was added to the bath.

Relaxation produced by oxmetidine was expressed as percentage of the effect induced by the ophylline and $-\log EC_{50}$ values were calculated.

Pulmonary airway resistance in vivo

Pulmonary airway resistance (R_{AW}) was determined according to the method of Advenier et al. (1983). Male guinea-pigs weighing 400-500 g were anaesthetized with urethane (1.25 g kg⁻¹) and small polyvinylchloride catheters were inserted into the left jugular vein and carotid artery. Intrathoracic pressure was measured by pleural puncture with a needle inserted into the pleural space through the sixth or seventh

intercostal space and connected through a catheter to a differential pressure transducer (Sanborn 2688). The animals were then placed in a body plethysmograph.

A Sanborn A 440 pneumotachometer was inserted through one of the plethysmograph walls thus allowing the measurement of air flow. Tidal volume was determined by electronic integration of the flow signal. Calculation of $R_{\rm AW}$ was made according to Amdur & Mead (1958). With this method, mean airway resistance is expressed as the ratio of transpulmonary pressure change to flow change occurring at two points of equal volume during inspiration and expiration. Since compliance is the same at these two points, this factor is eliminated from mean airway resistance calculations.

The experiments were performed on groups of at least 6 animals. An interval of 30 min elapsed before any drug administration. The actions of oxmetidine, cimetidine and ranitidine on the bronchoconstriction induced by histamine or acetylcholine (ACh) was tested on six groups of guinea-pigs, one group for each bronchoconstrictor and each H₂-receptor antagonist. Each animal received 3 or 4 intravenous doses of ACh $(25 \,\mu\mathrm{g \, kg^{-1}})$ or histamine $(20 \,\mu\mathrm{g \, kg^{-1}})$ at intervals of 5-10 min and the mean response was calculated. Oxmetidine, ranitidine and cimetidine were then administered intravenously and ACh or histamine was injected once more 10 min after each dose of antagonist. The results are expressed as percentage change of the initial increase in R_{AW} induced by the bronchoconstrictor agents. Under our experimental conditions, pulmonary airway resistance increased by 227 \pm 38% and $102 \pm 15\%$ after ACh and histamine, respectively. These values were not modified by prior administration of saline.

Statistical evaluation of data

Results obtained are presented as a mean \pm s.e.mean. Significant differences between the means were assessed by using one way analysis of variance. All the calculations were made according to Barlow (1983) by using a program running on an Apple IIe microcomputer.

Drugs

Oxmetidine (SKF 92994) and cimetidine were the generous gifts of Dr M.E. Parsons (Smith, Kline & French, Welwyn, U.K.); ranitidine hydrochloride was supplied by Laboratoires Glaxo (Paris, France); acetylcholine dihydrochloride and histamine hydrochloride were purchased from Laboratoires Lematte et Boinot (Paris, France) and Prolabo (Paris, France), respectively.

For the preparation of stock solutions, cimetidine was first dissolved in 0.1 NHCl and subsequently

adjusted to pH 6 with 0.1 N NaHCO₃. Ranitidine and oxmetidine were dissolved in distilled water. Dilutions were made with nutritive solutions or saline for *in vitro* and *in vivo* experiments, respectively.

Results

In vitro experiments

The effects of oxmetidine, ranitidine and cimetidine on guinea-pig isolated trachea are summarized in Figure 2. Oxmetidine was able to inhibit in a concentration-dependent fashion the contractions induced by ACh $(2 \times 10^{-5} \text{ M})$, histamine $(2 \times 10^{-5} \text{ M})$ and KCl $(1 \times 10^{-2} \text{ M})$, its $-\log \text{ EC}_{50}$ s being 3.46 ± 0.11 ,

 4.61 ± 0.09 and 4.20 ± 0.12 , respectively. Figure 3 depicts the results of a typical experiment in which histamine or KCl was employed as a contractile agent. In contrast to oxmetidine, the other H_2 -antagonists tested, namely ranitidine and cimetidine, were completely ineffective (Figure 2). Each of the three compounds, when added alone to the bath, was unable to modify the basal tone of the preparation.

When tested in Ca²⁺-free, K⁺-enriched medium, the inhibitory effect of oxmetidine on Ca²⁺-induced contractions was observed at concentrations close to those needed to counteract the spasmogenic effect of histamine in normal Krebs solution (Figure 4). In these experimental conditions, both cimetidine and ranitidine were again inactive.

The effect of oxmetidine on human bronchi precon-

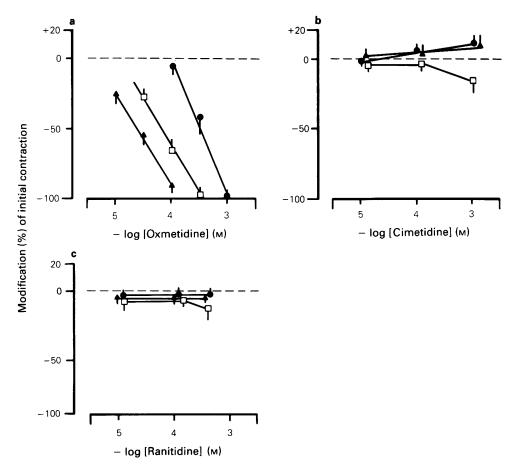


Figure 2 Effect of (a) exmetidine, (b) cimetidine and (c) ranitidine on acetylcholine $(2 \times 10^{-5} \,\mathrm{M}, \, \bullet)$, histamine $(2 \times 10^{-5} \,\mathrm{M}, \, \bullet)$ and KCl (I × 10⁻² M $\, \Box$)-induced contractions in guinea-pig isolated trachea. Each point represents the mean of the values obtained from 6-8 experiments. Vertical lines show s.e.mean.

tracted with ACh $(2 \times 10^{-5} \,\mathrm{M})$, histamine $(2 \times 10^{-5} \,\mathrm{M})$ or KCl $(1 \times 10^{-2} \,\mathrm{M})$ is depicted in Figure 5. In this preparation, too, the compound induced a concentration-dependent relaxation regardless of the excitatory stimulus. The calculated $-\log \,\mathrm{EC_{50}}$ values were $4.56 \pm 0.12, 4.55 \pm 0.09$ and 4.62 ± 0.05 against ACh, histamine and KCl, respectively.

In vivo experiments

In line with the *in vitro* experiments, oxmetidine was able to reduce significantly the increase in $R_{\rm AW}$ induced by both ACh and histamine in the anaesthetized guinea-pig (Table 1). Once again, ranitidine and cimetidine did not have any significant effects.

Discussion

Results of the present investigation show that, unlike ranitidine and cimetidine, oxmetidine was able to inhibit the contractions induced by the different test agents (ACh, histamine and KCl) on guinea-pig airways both *in vitro* and *in vivo*, as well as on human bronchi *in vitro*. The concentrations required to exert such a pharmacological action in the *in vitro* experiments were much higher than those shown to block H₂-receptors in guinea-pig atria and papillary muscle (Blakemore *et al.*, 1980; Coruzzi *et al.*, 1983a,b). These concentrations are, however, in the same range as those found by Bertaccini *et al.* (1983) and Black *et al.* (1985) to affect cardiac and vascular smooth muscle. In accordance with *in vitro* data, doses employed in *in vivo* experiments were greater than those required to inhibit histamine or pentagastrin-induced acid secretion (Blakemore *et al.*, 1980).

Besides the high doses required, two further observations suggest that this peculiar effect of oxmetidine is completely independent of H₂-receptor blockade: (1) the inactivity – under the same experimental conditions – of ranitidine and cimetidine, and (2) the knowledge that H₂-receptor

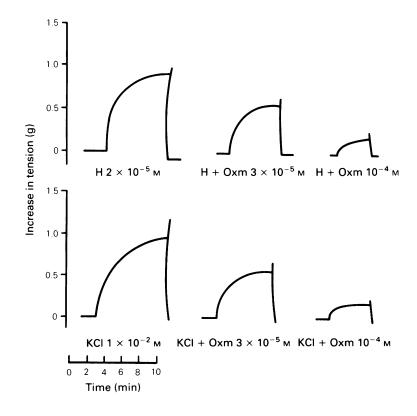


Figure 3 Guinea-pig isolated trachea: results of a typical experiment showing the inhibitory effect of oxmetidine (Oxm) on histamine $(H, 2 \times 10^{-5} M)$ and KCl $(1 \times 10^{-2} M)$ -induced contractions.

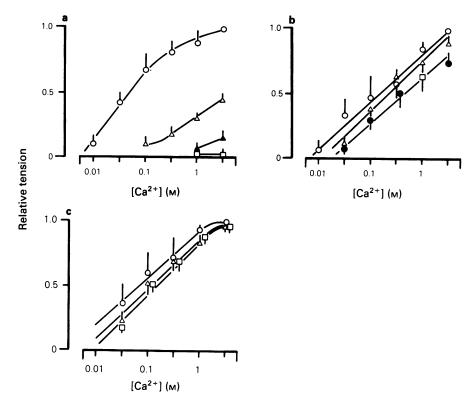


Figure 4 Guinea-pig tracheal spirals: effect of (a) oxmetidine, (b) cimetidine and (c) ranitidine on concentration-response curves to calcium. Responses were obtained in a potassium-rich (30 mm) solution in the absence (O) and presence of different concentrations (\bullet , 3×10^{-6} M; Δ , 3×10^{-5} M; Δ , 1×10^{-4} M; \Box , 3×10^{-4} M) of the antagonists. The contraction caused by CaCl₂ 3 mM in control experiments was arbitrarily taken as 1. Each point represents the mean of the values obtained from 4-6 preparations. Vertical lines show s.e.mean.

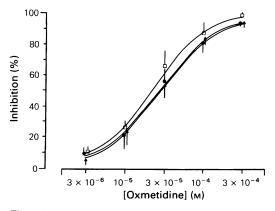


Figure 5 Inhibitory effect of oxmetidine against acetylcholine $(2 \times 10^{-5} \,\mathrm{M}, \, \bigcirc)$, histamine $(2 \times 10^{-5} \,\mathrm{M}, \, \triangle)$ and KCl $(1 \times 10^{-2} \,\mathrm{M}, \, \square)$ -induced contractions on human isolated bronchus preparation. Each point represents the mean of values obtained from 4–6 experiments. Vertical lines show s.e.mean.

stimulation is usually associated with bronchial smooth muscle relaxation. As a consequence, antagonists acting at H,-receptors usually enhance the response of guinea-pig tracheo-bronchial smooth muscle to histamine (for review see Eyre & Chand, 1982). Based on the results obtained on the rabbit isolated heart, Coruzzi et al. (1982) suggested that oxmetidine has a mechanism of action resembling that of the 'classical calcium channel antagonists', such as verapamil. Results of the present investigation, however, do not support this hypothesis. Indeed, on the guinea-pig isolated trachea as well as in the human bronchi, oxmetidine concentrations required to inhibit K⁺- or Ca²⁺-induced contractions are close to those necessary to counteract both ACh and histamineinduced contractions. It has been shown previously that, in these preparations, the 'classical' calcium channel antagonists (e.g. verapamil, diltiazem, nicarpidine) are able to inhibit K⁺-induced as well as Ca²⁺induced contractions at much lower concentrations than those required to inhibit the contractile effect of

 $+22 \pm 17$

Compound	3	Dose (mg kg	⁻¹) <i>30</i>
Acetylcholine			
Oxmetidine	$+32 \pm 27$	-30 ± 16	- 48 ± 15*
Cimetidine	-8 ± 28	$+62 \pm 41$	$+31 \pm 15$
Ranitidine	-8 ± 13	$+ 17 \pm 21$	$+ 19 \pm 16$
Histamine			
Oxmetidine	$+38 \pm 27$	$-26 \pm 6*$	$-53 \pm 14*$
Cimetidine	$+ 24 \pm 27$	$+ 18 \pm 22$	$+28 \pm 31$

Table 1 Effects of oxmetidine, cimetidine and ranitidine on the increase in pulmonary airway resistance R_{AW} induced in anaesthetized guinea-pigs by acetylcholine or histamine

Results, expressed as % change (increase (+), decrease (-)), are means \pm s.e.mean of the values obtained from 6-8 experiments.

+2921

 -31 ± 26

ACh and histamine (Advenier et al., 1984). On guineapig isolated trachea, as on the rabbit isolated aorta (Bertaccini et al., 1984), oxmetidine behaved as an antispasmogenic agent, its effect being independent of the type of stimulant employed to contract the preparation.

Ranitidine

The ability of oxmetidine to antagonize, to approximately the same extent, the stimulatory effect of compounds acting via different mechanisms (KCl exerts a direct action on tracheal smooth muscle and involves both depolarization and influx of Ca²⁺ (Foster et al., 1983), whereas ACh and histamine involve depolarization with little effect on Ca²⁺ influx (Ahmed et al., 1984) suggests that it may have a complex action, interfering with the transport at the membrane level and with the release or utilization of calcium ions at intracellular sites. Indeed, in the rabbit aorta oxmetidine behaved similarly to nitroglycerin, a compound which is assumed to act predominantly on intracellular calcium (Ito et al., 1980).

In our experimental conditions, neither ranitidine nor cimetidine modified the bronchoconstrictor effects of ACh and histamine either in vitro or in vivo. Our results are in accord with those of Duncan et al. (1980), who showed that, even at high doses, cimetidine and ranitidine were unable to produce a significant displacement of the concentration-response curves of airway smooth muscle preparations from guinea-pigs to contractile agents. However, Opkapo et al. (1978) found that histamine- but not ACh-induced contractions were greater in the

presence of metiamide, a former imidazole H₂-receptor antagonist. This potentiation has been attributed to the block of H₂-receptors mediating relaxation and was further confirmed by subsequent investigations involving the use of selective H₂-agonists, e.g. dimaprit or 4-methylhistamine (Eyre & Chand, 1982).

The lack of activity of ranitidine on ACh-induced contractions in guinea-pig trachea shows that in this tissue, unlike others, this H₂-antagonist is devoid of cholinomimetic action. Therefore, in agreement with the conclusions of Bertaccini (Bertaccini & Coruzzi, 1984b), the cholinergic-like effect of ranitidine is not ubiquitous and varies conspicuously between the different tissues and animal species.

Our results in vivo with the other two H₂-antagonists concord with the finding that metiamide and cimetidine are inactive against the antigen-induced allergic bronchospasm in guinea-pigs (Pare & Miller, 1980).

We concluded that, in contrast to ranitidine and cimetidine, oxmetidine displays non-specific antispasmogenic activity on airway smooth muscle both in vitro and in vivo. This action, which is independent of H₂-receptor blockade, represents a side-effect of the drug which may be connected with effects on the cellular influx of Ca²⁺ and the intracellular release or action of this ion.

This work was supported partially by a grant from M.P.I. (Rome). The help of Roberto Tramacere in the preparation of the manuscript is greatly appreciated.

References

ADVENIER, C., CERRINA, J., DUROUX, P., FLOCH, A., PRADEL, J. & RENIER, A. (1983). Sodium cromoglycate, verapamil and nicarpidine antagonism to leucotriene D₄

bronchoconstriction. Br. J. Pharmac., 78, 301-306. ADVENIER, C., CERRINA, J., DUROUX, P., FLOCH, A. & RENIER, A. (1984). Effects of five different organic

^{*}P < 0.05

- calcium antagonists on guinea-pig isolated trachea. Br. J. Pharmac., 82, 727-733.
- ADVENIER, C., NALINE, E. & RENIER, A. (1986). Effects of Bay K 8644 on contraction of the human isolated bronchus and guinea-pig isolated trachea. *Br. J. Pharmac.*, 88, 33-39.
- AHMED, F., FOSTER, R.W., SMALL, R.C. & WESTON, A.H. (1984). Some features of the spasmogenic actions of acetylcholine and histamine in guinea-pig isolated trachealis. Br. J. Pharmac., 83, 227-233.
- AMDUR, M.O. & MEAD, J. (1958). Mechanics of respiration in unanaesthetized guinea-pigs. Am. J. Physiol., 192, 364-368
- BARLOW, R.B. (1983). Biodata Handling With Microcomputers. Cambridge: Elsevier/Biosoft.
- BERTACCINI, G. & CORUZZI, G. (1982). Cholinergic-like effects of the new histamine H₂-receptor antagonist ranitidine. *Agents Actions*, 12, 168-171.
- BERTACCINI, G. & CORUZZI, G. (1984a). H₂-receptor antagonists: side-effects and adverse effects. *Ital. J. Gastroenterol.*, **16**, 119–125.
- BERTACCINI, G. & CORUZZI, G. (1984b). Interaction between histamine H₂-receptor antagonists and the cholinergic system. In *Update: H₂ Receptor Antagonists* ed. Cohen, S. pp. 9–19, New York: Biomedical Information Corporation.
- BERTACCINI, G. & DOBRILLA, G. (1980). Histamine H₂-receptor antagonists: old and new generation. Pharmacology and clinical use. *Ital. J. Gastroenterol.* 12, 309-314
- BERTACCINI, G., MOLINA, E., BOBBIO, P. & FOGGI, E. (1981). Ranitidine increases lower esophageal sphincter pressure in man. *Ital. J. Gastroenterol.*, 13, 149-150.
- BERTACCINI, G., POLI, G., ADAMI, M. & CORUZZI, G. (1983). Effect of some new H₂-receptor antagonists on gastrointestinal motility. *Agents Actions*, 13, 157-162.
- BERTACCINI, G., POLI, E. & RADDINO, R. (1984). Action of oxmetidine on the rabbit aorta: comparision with some calcium entry blockers and nitroglycerin. *Pharmac. Res. Commun.*, 16, 161-170.
- BERTACCINI, G. & SCARPIGNATO, C. (1982). Histamine H₂-antagonists modify gastric emptying in the rat. *Br. J. Pharmac.*, 77, 443-448.
- BLACK, J.W., LEFF, P. & SHANKLEY, N.P. (1985). Further analysis of anomalous pK_B values for histamine H₂receptor antagonists on the mouse isolated stomach assay. Br. J. Pharmac., 86, 581-587.
- BLAKEMORE, R.C., BROWN, T.H., DURANT, G.J., EMMETT, J.C., GANELLIN, C.R., PARSONS, M.E. & RASMUSSEN, A.C. (1980). SKF 92994: a new histamine H₂-receptor antagonist. *Br. J. Pharmac.*, 70, 105P.
- BRINK, G., GRIMAUD, C., GUILLOT, C. & OREHEK, J. (1980). The interaction between indomethacin and contractile agents on human isolated airway muscle. *Br. J. Pharmac.*, 69, 383-388.
- CORUZZI, G., DI LISA, F., FERRARI, R., RADDINO, R. & BERTACCINI, G. (1982). Effect of the new H₂-receptor

- antagonist, oxmetidine, on the isolated rabbit heart. In *Advances in Studies on Heart Metabolism*. ed. Caldarera, C.M. & Harris, P. pp. 151-158, Bologna: CLEUB.
- CORUZZI, G., POLI, E. & BERTACCINI, G. (1983a). Cardiac effects of the new H₂-receptor antagonists. *Agents Actions*, 13, 173–178.
- CORUZZI, G., POLI, E., FESANI, F., MEDICI, D. & BERTAC-CINI, G. (1983b). Negative inotropic effect of some H₂receptor antagonists on the isolated human atria. Experientia, 39, 1284-1286.
- DAL NEGRO, R.W., TURCO, P. & ZOCCATELLI, O. (1984). Effects of orally administered cimetidine, ranitidine and oxmetidine on bronchomotor tone in normal volunteers. *Int. J. clin. Pharm, Res.*, 4, 481-482.
- DUNCAN, P.G., BRINK, C., ADOLPHSON, R.L. & DOUGLAS, J.S. (1980). Cyclic nucleotides and contraction/relaxation in airway muscle: H₁ and H₂ agonists and antagonists. J. Pharmac. exp. Ther., 215, 4343-442.
- EYRE, P. & CHAND, N. (1982). Histamine receptor mechanisms of the lung. In *Pharmacology of Histamine Receptors*. ed. Ganellin, C.R. & Parsons, M.E., pp. 298-322. Bristol: Wright.
- FOSTER, R.W., SMALL, R.C. & WESTON, A.H. (1983). The spasmogenic action of potassium chloride in guinea-pig trachealis. *Br. J. Pharmac.*, **80**, 553-559.
- GHELANI, A.M., HOLROYDE, M.C. & SHEARD, P. (1980). Response of human isolated bronchial and lung parenchymal strips to SRS-A and other mediators of asthmatic bronchospasm. *Br. J. Pharmac.*, 71, 107-112.
- GNASSOUNOU, J.P., ADVENIER, C. & SCARPIGNATO, C. (1986). Effect bronchodilatateur de l'oxmetidine. J. Pharmac. (Paris), 17, 437.
- GODFRAIND, T., KABA, A. & POLSTER, P. (1968). Differences in sensitivity of arterial smooth muscles to inhibition of their contractile response to depolarization by potassium. Arch. int. Pharmacodyn., 172, 235-239.
- GUILLOT, C., FORNARIS, M., BADIER, M. & OREHEK, J. (1984). Spontaneous and provoked resistance to isoproterenol in isolated human bronchi. J. Allergy clin. Immunol., 74, 713-718.
- ITO, Y., KITAMURA, K. & KURIYAMA, H. (1980). Action of nitroglycerine on the membrane and mechanical properties of smooth muscles of the coronary artery of the pig. Br. J. Pharmac., 70, 197-204.
- OKPAKO, D.T., CHAND, N. & EYRE, P. (1978). The presence of inhibitory histamine H₂-receptors in guinea-pig tracheobronchial muscle. J. Pharm. Pharmac., 30, 181-182.
- PARE, P. & MILLER, D. (1980). The effect of H₂-blocking agents on systemic anaphylaxis in guinea-pigs. *Ann. Rev. resp. Dis.*, 121, 749-752.
- SCARPIGNATO, C., BERTACCINI, G., ZIMBARO, G. & VITULO, F. (1982). Ranitidine delays gastric emptying of solids in man. Br. J. clin. Pharmac., 13, 252-253.
- VINCENC, K., HWANG, O. & VAN BREEMEN, C. (1984). Relaxation and contraction responses to histamine in the human lung parenchymal strip. Eur. J. Pharmac., 98, 201-210.

(Received July 7, 1986. Revised October 24, 1986.) Accepted November 1, 1986.)